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University of Patras, Patras, Greece. Fertile F₁ males and females from crosses between **Drosophila mauritiana** females and **D.melano**gaster or **D.simulans** males. It is known that in crosses between the sibling species **Drosophila simulans** and **D.mauritiana** (a species originated from **D.simulans**; Tsacas et al. 1981) fertile females and sterile males are yielded in the F_1 generation, independently of what species is used as female. Moreover, it is also known that in crosses between the sibling species **D.melanogaster** and

D.mauritiana, sterile females or males (in respect with the melanogaster sex used) are yielded in the F₁ generation. In spite of this general consensus, we present now data showing that fertile males and females can be obtained from some crosses between the sibling species pair mentioned above. Thus, in one out of 55 pair matings, where **mauritiana** was used as female and **melanogaster** as male, fertile F₁ males and females appeared. The fertility of these individuals was tested by mating them and noticing if F₂ (or F_n) progeny are yielded. These interspecific fertile hybrids have been named mame and are kept with success in our laboratory as a stock for 10 generations so far. In the opposite cross (? **melanogaster** x **mauritiana** σ), seven out of 43 pair matings were found to yield hybrid females (named mema) which are sterile when crossed with **melanogaster** or **mauritiana** males. It must be noted that as **melanogaster** we used the CyL⁴/Pm stock, while in a previous effort using another **melanogaster** stock (homozygous for the malate dehydrogenase fast form), we failed to get one successful mating (yielded offspring) out of 92 performed.

In another interspecific cross-type where mauritiana was used as female and simulans as male, two out of 82 pair matings yielded fertile F₁ males and females. These interspecific fertile hybrids have been named <u>masi</u> and are kept in our lab as a stock, like <u>mame</u>. In the case where mauritiana is used as male and simulans as female, 63.41% (26/41) successful matings obtained. Each such successful pair mating yielded females and males which are sterile when crossed with each other but fertile when crossed with mauritiana (males) and simulans (males or females). When mauritiana females are crossed with the above described interspecific hybrids (named sima), no progeny are produced. The same is also true in the cross

Table 1. Successful and unsuccessful pair-matings between pairs of D.melanogaster (mel), D.simulans (sim) and D.mauritiana (maur), as well as between fertile interspecific hybrids* and their parents.

species pair	No. of matings successful unsuccessful total			
♀ sim x maur ♂	26	15	41	
♀ maur x mel ď	1	54	55	
♀ maur x sim ď	2	80	82	
♀ mel x maur ď		36	43	
♀ masi x sim ♂	15	27	42	
9 masi x maur d	16	1	17	
9 maur x masi d	0	49	49	
♀ sim x <u>masi</u> ♂	1		4	
♀ mame x mel ď	13	0	13	
a mame x maur d	• 1	7	8	
9 maur x mame d	0	38	38	
♀ mel x mame ď	8	0	8	
♀ mema x mel ď	0	14	14	
9 mema x maur d	0	28	28	
♀ sima x sim ♂	7	3	10	
♀ sima x maur ď	3	3	6	
♀ maur x sima ď	0	16	16	
♀ sim x sima ♂	9	0	9	
♀ masi x mame ď	5	18	23	
♀ mame × masi ♂	7	25	32	

*The name of the interspecific hybrids is composed of two syllables. The first is from the \mathfrak{P} and the seond from the σ species-parent name. ⁹ mauritiana x mame σ . In crosses ⁹ mame x masi σ (and the reciprocal), some pair matings are successful, yielding females and males which possibly get genes from three different species (melanogaster, simulans, mauritiana). In Table 1 we show all the above information described and in Table 2 we give a brief summary of the reproductive isolation status among the three sibling species examined as it was found in our lab (with the strains of the species we used).

It must be noted that the combination **melanogaster-simulans** was tested and our findings verify the previous consensus (see Table 2), that is, sterile F_1 females or males are produced, dependent on the **melanogaster** sex used. However, in a mass cross (6 § **melanogaster** x **simulans** σ 6) a part of the sterile females obtained as expected, 3 hybrid males were also

Table 2. Hybridization possibilities betweenthe 3 sibling species, melanogaster, simulans,mauritiana, as it has been found in our lab.

parents	Ŷ	ď	
♀ mel x sim ď	sterile		
♀ sim x mel ď		sterile	
♀ mel x maur d	sterile		
♀ maur x mel ď	fertile	fertile	
♀ sim x maur ď	fertile*	fertile*	
♀ maur x sim ♂	fertile	fertile	

*but sterile when crossed with each other (see text)

Table 3. Mating propensities in multiple choice experiments involving the interspecific fertile hybrids <u>masi</u> and its parental species, mauritiana and simulans.

Cross A x B	₽AxAo*	₽AxBo	\$BxA♂	₽BxBơ		Sexual Isolation Index ± S.E.
sim x masi	25	22	13	27	6	0.195 ± 0.105
maur x masi	5	14	15	37	8	0.183 ± 0.116

* in each chamber existed 12 virgin females and 12 males from each stock.

noticed. They electrophorized for the alcohol dehydrogenase and exhibited an intermediate (between melanogaster and simulans) electrophoretic pattern.

The situation where mauritiana females do not give offsprings with masi or mame males, was further investigated in an attempt to see if this situ-

ation is due to pre- or post-mating isolation mechanism. Thus, performing multiple choice experiments (see for methodology Kilias & Alahiotis 1982), the mating propensities of our interspecific hybrids with their parental species were determined. As it is shown in Table 3, mauritiana females copulate with masi males, a fact which demonstrates that the absolute reproductive isolation observed between them is not due to premating mechanisms. Furthermore the same table shows that no significant sexual isolation toward homogametic mating has been developed. Taking into consideration the melanogaster-mame combination, we observed in preliminary experiments, that all the four possible mating types can be obtained, while this is not true in the case of mauritiana-mame combination where mauritiana females do not seem to copulate with mame males.

The situation described here regarding the reproductive isolation status between three sibling species in the **D.melanogaster** subgroup differs with that which it was so far known. These differences may be based on the genetic composition of our strains used (e.g. CyL⁴/Pm for **D.melanogaster**; **D.simulans** was captured recently from a Greek natural population) or on some evolutionary changes of **D.mauritiana** under the laboratory conditions where lately is maintained.

The implication of these hybrids to the study of the speciation mechanisms is obvious and can be proved important in understanding the evolution of interspecific reproductive isolation. The elucidation of the detailed genetic organization of these hybrids (the status of which was also verified by electrophoretic and cytogenetic criteria) will contribute greatly to the approach of the above purpose.

References: Kilias, G. & S.N. Alahiotis 1982, Evolution 36:121-131; Tsacas et al. 1981, in: The Genetics and Biology of Drosophila, Ashburner, Carson & Thompson (eds.), Acd. London, Vol 3a:197-259.

Albers, K.B.M. and R. Bijlsma. University of Groningen, Haren, Netherlands. Selection for increase in tolerance with respect to xenobiotics in Drosophila melanogaster. It is well known that **D.melanogaster** can readily become tolerant for a wide range of toxic chemicals when present in its environment. To obtain insight in the process of developing tolerance, two different populations of **D.melanogaster** were subjected to selection on five xenobiotics. Selection was

performed by rearing the flies in cages on food supplemented with the xenobiotics. During the experiment the concentrations were increased regularly in 7-8 steps and the initial concentrations and the concentrations after 17 months (the moment the tolerance levels were determined) of the different chemicals are given: (1) phenobarbital (sodium salt): $250 \div 1300$ ppm; (2) rotenon (a commercial anti-flea powder containing 0.9% rotenon was used): $6 \div 24$ ppm; (3) malathion (a commercial preparation containing 50% malathion was used): $0.15 \div 0.65$ ppm; (4) carbaryl (also a commercial preparation containing 50% carbaryl was used): $6 \div 28$ ppm; (5) DDT: $15 \div 65$ ppm. The first chemical is used as a drug; the other four are or have been used as insecticides.

For the experiments two different sets of each six population cages were established. The Bogota populations were initiated with 36 independent lines isolated from the Bogota base population as described by Bijlsma (1980). The 50 x 50 populations were initiated with 40 lines from the second reisolation from the original 50 x 50 base population as described by Bijlsma & Van Delden (1977). All cages of each set were provided with standard food (for description of the food see Bijlsma 1980) for the first weeks to get them well established. Thereafter one cage of each set was kept on this food (control) while the others were supplied with standard food supplemented with one of the five toxic compounds. To standardize the selection pressures somewhat, the initial concentrations were chosen in such a way that the larval viability was approximately 40-60%; as a results the density in the cages was kept well over a thousand individuals. When the concentrations were increased during the experimental period, it was also ensured that the population density stayed above this level.